

Title Polyphasic characterization of antibiotic resistant and virulent enterococci isolated from animal feed and stored-product insects

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Abstract

Feed samples and live stored-product insects from feed mills and swine farms were collected and cultured for *Enterococcus* spp. The mean concentration of enterococci in insect and feed were $2.7 \pm 0.5 \times 10^1$ cfu/insect and $6.3 \pm 0.7 \times 10^3$ cfu/g respectively. A total of 362 isolates of enterococci collected from 89 feed samples and 228 stored-product insects were identified to the species level using PCR. These isolates were represented by *Enterococcus casseliflavus* (53.0%), *E. gallinarum* (20.4%), *E. faecium* (16.2%), *E. hirae* (5.2%), and *E. faecalis* (5.0%). Enterococci were phenotypically resistant to tetracycline (48.0%), erythromycin (14.3%), streptomycin (16.8%), kanamycin (12.1%), ciprofloxacin (11.0%), ampicillin (3.3%), and chloramphenicol (1.1%). All isolates were susceptible to vancomycin and gentamicin. Tetracycline resistance was encoded by *tetM* (50.0%), *tetO* (15.1%), *tetK* (0.5%), *tetS* (0.2%) and other unknown tetracycline determinants. Enterococci carried virulence genes including gelatinase (*gelE*; 21.5%), an enterococcus surface protein (*esp*; 1.9%), and cytolysin (*cylA*; 2.2%). An aggregation substance (*asaI*) gene was detected in 61.0% of *E. faecalis* isolates. Fifty percent of *E. faecalis* isolates were phenotypically tested positive for aggregation substances. Enterococci with *cylA* genes were hemolytic (52.0%) and with *gelE* genes were gelatinolytic (18.5%). The *ermB* gene, encoding erythromycin resistance was detected in 8.8% of the total isolates. The Tn916/1545 family of conjugative transposons was detected in 10.7% of the isolates.

Laboratory experiments showed that adults of the red flour beetle, *Tribolium castaneum* (Herbst), fed on poultry and cattle feeds inoculated with *E. faecalis* OG1RF:pCF10, were able to successfully acquire enterococci and contaminate sterile poultry and cattle feeds. To assess the potential of horizontal gene transfer, conjugation assays were carried out with *E. faecalis* using a donor (wild strains) and recipient (*E. faecalis* OG1SSP) in ratio of 1:10. Only one isolate (1 out of 18 *E. faecalis*) could transfer *tetM* to a recipient using broth mating. However, filter mating assay, followed by PCR confirmation

revealed that 89.0% (16 out of 18 *E. faecalis*) of isolates could transfer *tetM* to *E. faecalis* . Transfer ratios of transconjugant per recipients ranged from 2.6×10^{-4} to 1×10^{-9} .

In summary, feed (52.0%) and stored-product insects (41.6%) collected from feed mills and swine farms carried antibiotic-resistant and potentially virulent enterococci. Our study showed that *T. castaneum* , a pest commonly associated with feed, served as a potential vector for enterococci in the feed environment. Conjugation assays followed by PCR confirmed presence of the *tetM* gene on a mobile genetic element(s) such as Tn *916* and may be horizontally transferred to other *Enterococcus* species and to other bacteria of clinical significance.